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MECHANISM OF BICARBONATE ACTION ON PHOTOSYNTHETIC ELECTRON TRANSPORT IN BROKEN CHLOROPLASTS

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In CO₂-depleted chloroplasts electron transport between the Photosystem II electron acceptor Q and plastoquinone is largely suppressed. In the presence of a high concentration of sodium formate (greater than 10 mM), which probably binds to the bicarbonate site, addition of bicarbonate restores the ferricyanide Hill reaction only after incubation in the dark. With lower formate concentrations bicarbonate is able to restore electron transport in the light. The Hill reaction rate in CO₂-depleted chloroplasts after bicarbonate addition, divided by the rate in CO₂-depleted chloroplasts before bicarbonate addition, shows a sharp optimum at pH 6.5. Furthermore, the rate-limiting step in bicarbonate action is probably diffusion. The results are explained in terms of a hypothetical model: the bicarbonate-binding site is located at the outer side of the thylakoid membrane, but not directly accessible from the 'bulk'. To reach the site from the bulk, the molecule has to pass a channel with negatively charged groups on its side walls. In the light these groups are more negatively charged than in the dark. Therefore, the formate ion cannot exchange for bicarbonate in the light, and a dark period is necessary to enable exchange of formate for bicarbonate.

Introduction

 ${\rm CO_2}$ is not only required as a substrate for ribulosebisphosphate carboxylase, but is also indispensable for photosynthetic electron transport [1]. A major site of action is between the Photosystem II electron acceptor Q and plastoquinone [2-4]. (For a review see Ref. 5.)

It was not known whether CO₂ or HCO₃ is the active species regulating electron flow until Good [6] presented evidence that HCO₃ is involved in the bicarbonate effect. He showed that it was easier to lower the rate of the Hill reaction with ferri-

cyanide by CO_2 depletion in the presence of HCO_3^- -like anions (e.g., HCO_2^- , $CH_3CO_2^-$) than in their absence. These anions would displace HCO_3^- from the binding site, causing a lowering of the Hill reaction rate, since binding of HCO_3^- (or CO_2) is required for photosynthetic electron flow.

The mode of action of HCO_3^- is not yet understood, although it has been suggested that HCO_3^- forms a complex with the two-electron carrier R (or B) in the dark [7].

The ratio of the Hill reaction rate after the addition of HCO₃ to that in CO₂-depleted chloroplasts has been shown to be strongly dependent on pH. At pH 5.8 the restoration of the Hill reaction in CO₂-depleted chloroplasts requires more HCO₃ addition than at pH 6.8 [8]. Furthermore, a maximal effect has been observed at pH 6.5-7.0. At pH 8.0-8.5 the ratio of the Hill reaction rate of

^{*} To whom correspondence should be addressed. Abbreviations: DCMU, 3-(3,4-dichlorophenyl)-1,1-dimethylurea; Mes, 4-morpholineethanesulfonic acid; simeton, 2-methoxy-4,6-bis(ethylamino)-1,3,5-triazine; Chl, chlorophyll.

+HCO $_3^-$ /-HCO $_3^-$ is nearly 1.0 [3]. In this pH range the concentration of HCO $_3^-$ is maximal in comparison to the concentrations of CO $_2$ and CO $_3^-$ (p K_a of H $_2$ CO $_3$ is 6.4; p K_a of HCO $_3^-$ is 10.2 at 25°C). This suggests that HCO $_3^-$ is not the only important factor required in photosynthetic electron transport.

In CO₂-depleted chloroplasts, suspended in media containing 100 mM sodium formate, the Hill reaction has been shown to be restored by adding HCO₃ followed by dark incubation for 1-2 min. Under these conditions no restoration is possible in the light [7,8]. However, we have been able to observe reactivation of the Hill reaction rate by addition of HCO3 to CO₂-depleted chloroplasts in HCO₂-free media in the light. We present evidence based on this reactivation in the light and on the dependence of the bicarbonate effect on pH and on temperature that HCO₃ and CO₂ are both involved in 'bicarbonate action'. We suggest that HCO₃ is the species that binds at the thylakoid membrane, but that the molecule reaches its binding site mainly in CO₂ form. The HCO3-binding site is covered by the negatively charged membrane and therefore is not readily accessible for HCO3 from the bulk. However, the uncharged CO₂ can pass easily. The binding site is not specific for HCO₃. Other organic anions like HCO₂, CH₃CO₂ and the herbicide, 4,6-dinitro-ocresol, can also bind to the site, but only after binding of HCO₃ is electron transport possible.

A preliminary report of this work has been presented earlier [9].

Materials and Methods

Chloroplasts were isolated from 10-day-old peas (*Pisum sativum*, cv. Rondo) and broken, as described elsewhere [10]. The broken chloroplasts were suspended in a standard medium, containing 50 mM sodium phosphate, 100 mM NaCl, 100 mM sodium formate and 5 mM MgCl₂ (pH 5.0); chlorophyll concentration was 50 μ g/ml. The suspension was shaken vigorously for 10 min at room temperature in the dark, while N₂ gas was bubbled through. The CO₂-depleted chloroplasts were pelleted and resuspended in the standard medium at pH 6.5, unless indicated otherwise. All centrifuge tubes and media were made CO₂-free before use.

Electron transport was measured as O2 produc-

tion as described earlier [11], using 0.5 mM ferricyanide as electron acceptor.

When HCO₂-free CO₂-depleted chloroplasts were needed, the chloroplasts were washed after CO₂ depletion and resuspended in a medium containing 50 mM Mes, 0.4 M mannitol and 5 mM MgCl₂ (pH 6.5).

Results

Reactivation by bicarbonate in the light and in the dark. We have been able to obtain CO₂-depleted chloroplasts with a ferricyanide Hill reaction rate of less than 0.5 μ mol O₂/mg Chl per h at pH 6.5 in the standard medium. Fig. 1 shows that maximal reactivation of the Hill reaction rate is observed after the adddition of 10 mM HCO₃ (which does not cause a detectable pH change) and 2 min dark incubation. Then the ferricyanide Hill reaction is 20–30 μ mol O₂/mg Chl per h. Half reactivation occurs at 1 mM HCO₃. Fig. 1 confirms the result obtained earlier by Khanna et al. [3]. Prolongation of the dark incubation period does not cause a further increase of the Hill reaction rate. Control chloroplasts, i.e., not CO2-depleted and reactivated, show an uncoupled electron flow rate with ferricyanide in the standard medium of about 150 µmol O₂/mg Chl per h. The CO2-depleted chloroplasts also appear to be

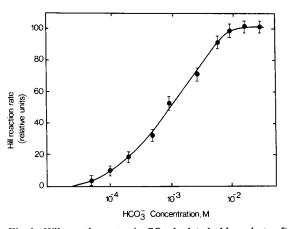


Fig. 1. Hill reaction rates in CO_2 -depleted chloroplasts after 2-min dark incubation with various amounts of HCO_3^- . The measurements were performed in the standard medium at pH 6.5. 100 relative units are equivalent to 21.5 μ mol O_2/mg Chl per h.

uncoupled, presumably due to the high salt concentration of the depletion and reaction medium [3].

The site which is irreversibly damaged by the depletion procedure appears to be located at or before the reoxidation of Q-, because after trypsin treatment, which is known to make Q accessible for ferricyanide [12-14], or using silicomolybdate as an electron acceptor, CO2-depleted chloroplasts yield a rate of the Hill reaction not very different from the rate in the presence of saturating amounts of HCO₃ [3,15]. Addition of exogenous Photosystem II donors, such as 2 mM diphenylcarbazide or 70 µM benzidine, does not increase the dichlorophenolindophenol Hill reaction rate in CO2-depleted chloroplasts to which HCO₃ is added (Vermaas, W.F.J. and Van Rensen, J.J.S., unpublished observations), indicating that the site of the irreversible damage is situated after (or at) rereduction of Z⁺. Therefore, we suggest that the irreversible damage is located between rereduction of Z⁺ and reoxidation of Q⁻.

In a completely different system, i.e., non-HCO₃-depleted chloroplasts, Crane and Barr [16] found that addition of HCO₃ inhibited the DCMU-insensitive silicomolybdate reduction by Photosystem II. (For a discussion of this phenomenon, see Ref. 5.)

In the standard medium, no increase of the Hill reaction rate is observed in CO₂-depleted chloroplasts after the addition of HCO₃ in the light. A dark incubation period seems to be necessary, in agreement with Stemler [7]. However, in the absence of high concentrations of HCO₂, HCO₃ is able to increase the Hill reaction rate in CO₂-depleted chloroplasts also in the light (Fig. 2). Addition of the other constituents of the standard medium (100 mM NaCl or 50 mM sodium phosphate, pH 6.5) does not prohibit reactivation in the light, only HCO₂ does.

The CO_2 -depleted chloroplasts suspended in the HCO_2^- -free medium show a measurable O_2 production rate, presumably caused by the absence of the HCO_3^- -binding inhibitor, HCO_2^- . The minimal O_2 production rate under these conditions is about 5 μ mol O_2/mg Chl per h. After the addition of HCO_3^- , the ferricyanide Hill reaction rate is stimulated by a factor of 3-6.

Action of formate. It has been suggested that HCO_2^- binds to the same site as HCO_3^- does [3,6]. However, electron flow occurs only when HCO_3^-

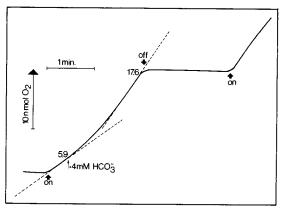
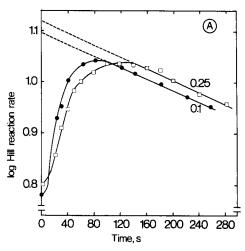


Fig. 2. Reactivation of CO₂-depleted chloroplasts by addition of 4 mM HCO₃ in the light. Reaction medium: 4 mM Mes, 0.4 M mannitol, 5 mM MgCl₂, 0.5 mM ferricyanide and Chl concentration of 33 μ g/ml; pH was 6.5. Numbers along trace: μ mol O₂/mg Chl per h.

is bound to the site. To quantify the formate effect on HCO₃ action, different HCO₂ concentrations were added to CO2-depleted HCO2-free chloroplasts. In the light, at time zero, 10 mM HCO₃ was added and the reactivation as a function of time was measured (Fig. 3A). An initial increase in the ferricyanide Hill reaction is observed after the addition of HCO3, followed by a decrease due to deactivation of the chloroplasts (e.g., by denaturation of proteins). When the Hill reaction rate is plotted on a logarithmic scale against the time on a linear scale, this deactivation appears to give a straight line. So, $d = k \cdot e^{-ct}$, in which d is deactivation, c and k are constants and t is time. Deactivation appears to be a (pseudo) first-order reaction. We assume that deactivation starts at time zero and follows a straight baseline. Now the differences between the baselines and the observed values were calculated as $\%\Delta\log(\text{Hill reaction rate})$; this is the percentage of the difference between the baseline and observed values, both taken as their logarithms. The difference at time zero was taken as 100%. These values were plotted against time. Fig. 3B shows the results for various HCO₂ concentrations. From this figure, it is easily seen that even 0.1 mM HCO₂ has an appreciable effect in decelaration of bicarbonate action in the light. No exact quantitative data can be obtained from this figure, because it is not known how much HCO2 is left in the sample after washing and resuspension in HCO₂-free medium.



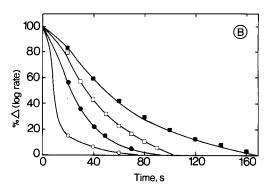


Fig. 3. (A) Reactivation of the Hill reaction in the light by addition of 10 mM HCO_3 at time zero in the presence of $0.1 \text{ mM } (\bullet)$ or $0.25 \text{ mM } (\Box) \text{ HCO}_2$. Reaction medium: 50 mM Mes, 0.4 M mannitol, 5 mM MgCl_2 , 0.5 mM ferricyanide, $33 \mu \text{g Chl/ml}$ and pH 6.5. (B) Relative difference between baseline and measured values as shown in A. \circ , without HCO $_2$; \bullet , 0.1 mM; \Box , 0.25 mM; \blacksquare , 10 mM HCO_2 added. For explanation, see text.

From this plot we conclude that one cannot expect a measurable reactivation by adding HCO_3^- in the light to CO_2 -depleted chloroplasts when 100 mM HCO_2^- is present, because in the presence of 10 mM HCO_2^- the reactivation is already very slow.

The dependence on pH. As mentioned in the Introduction, the ratio of the Hill reaction rate after the addition of HCO_3^- to that in its absence is strongly dependent on pH. To obtain a more precise value for the influence of pH on this ratio than that measured earlier [3], we have prepared partially CO_2 -depleted chloroplasts. Their ferricyanide Hill reaction rate was about 3 μ mol O_2 /mg Chl per h at pH 6.5. These chloroplasts were suspended in CO_2 -free media at a different pH, and the reactivation by addition of 10 mM HCO_3^- was measured (Fig. 4).

We have found a sharp pH optimum at pH 6.5. This is close to the pK_a of H_2CO_3 , which is 6.37 at 25°C. Our result is a good indication that both HCO_3^- and H_2CO_3 or CO_2 are involved in the reactivation of CO_2 -depleted chloroplasts. However, we have to realize that the pH very close to the negatively charged membrane surface may be lower, due to the ζ -potential [17].

The dependence on temperature. At 25°C, 2 min dark incubation of CO₂-depleted chloroplasts in standard medium with HCO₃ is just enough for maximal restoration of electron transport.

We have investigated the influence of temperature on bicarbonate action by calculating the Q_{10} value (i.e., the Hill reaction rate at $T = t^{\circ}C$ over that at $T = (t - 10)^{\circ}C$). For that purpose the ferricyanide Hill reaction rate in CO₂-depleted chloroplasts was measured after 2 min dark incubation with 1 or 10 mM HCO₃ at different temperatures. From Fig. 5, a Q_{10} value of 2.0 ± 0.2 can be calculated ($0 \le T < 25^{\circ}C$). This is close to the value of 1.8 obtained

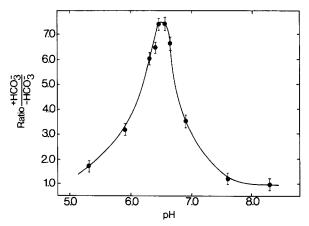


Fig. 4. Ratio of the Hill reaction rate in the presence of 10 mM HCO_3 divided by that in its absence in CO_2 -depleted chloroplasts at various pH values. Measurements were performed in the standard medium at various pH values.

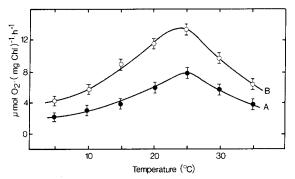


Fig. 5. Hill reaction rates in CO_2 -depleted chloroplasts after 2-min incubation with 1 mM (A) or 10 mM (B) HCO_3^- at various temperatures. The measurements were performed in standard medium. Since the chloroplasts were very thoroughly depleted, the Hill reaction rates in the presence of HCO_3^- are rather low.

in normal (not CO_2 -depleted) chloroplasts, suspended in the same standard medium (unpublished results). Because the Q_{10} values of 2.0 and 1.8 are almost the same, we conclude that bicarbonate action per se is quite insensitive to temperature. A low Q_{10} value might point to diffusion or to an enzymatic reaction as the rate-limiting step in bicarbonate action.

If an enzymatic reaction is involved in this rate-limiting step, it would probably be a carbonic anhydrase-catalyzed reaction. Therefore, we have investigated the influence of carbonic anhydrase on the bicarbonate reactivation of CO₂-depleted chloroplasts in the light in a medium containing little HCO₂. We have found no difference in the reactivation rate with or without carbonic anhydrase. For this reason, it is very probable that diffusion is the rate-limiting step in bicarbonate action. Sarojini and Govindjee (personal communication) have shown that carbonic anhydrase affects equilibrium at 5°C, but carbonic anhydrase has no effect at 25°C as we have found.

Discussion

Depleting broken chloroplasts of CO_2 causes a complete inhibition of electron transport. Although there is definitely a major effect on the reducing side of Photosystem II [2-5], which is the suppression of electron flow between the electron acceptor Q and

plastoquinone in the absence of HCO_3^- , it has been argued that there should be a CO_2 effect on 'watersplitting', because CO_2 might be the immediate source of photosynthetically evolved O_2 [18–20]. We discuss our results in relation to the effects of CO_2 and HCO_3^- at the reducing side of Photosystem II

The reactivation by HCO₃ is affected by HCO₂ [9,20-22]. We observed that in the presence of 100 mM HCO₂, half-maximal reactivation in CO₂-depleted chloroplasts is observed by incubation in the dark for 2 min in the presence of 1 mM HCO₃ (Fig. 1). At the pH of the medium (6.5) nearly all added formate is in HCO_2^- form, since the p K_a of formate is 3.7. At pH 6.5 about half of the added HCO₃ is in HCO₃ form, the other half in the CO₂ form. This means that at half reactivation at pH 6.5, 0.5 mM HCO₃ and 100 mM HCO₂ are present. Thus, the dissociation constant of the HCO₂-binding site complex is about 200-times higher than the dissociation constant of the HCO₃-binding site complex, assuming that there is no cooperativity. Fig. 3B shows that the presence of 0.1 mM HCO₂ already has a significant effect on bicarbonate action in the light. This would suggest that in fully CO₂-depleted chloroplasts in a medium without HCO_2^- , even 0.5 μM HCO_3^- will show a measurable effect on the Hill reaction rate (0.1 mM divided by 200). This might be the reason why it is almost impossible to prepare CO₂-depleted chloroplasts in media which do not contain HCO₂. Also, Khanna et al. [3] have observed that a comparable restoration of electron transport in CO2-depleted chloroplasts occurs at lower concentrations of HCO₃ in the absence than in the presence of 100 mM HCO_2^- .

We suggest that the \(\xi\)-potential, defined as the electrokinetic potential at the hydrodynamic plane of shear [17], plays an important role in the bicarbonate effect. The \(\xi\)-potential is more negative in the light than in the dark [23]. Figs. 2 and 3 show that HCO\(\frac{3}\) can restore the Hill reaction in CO\(\frac{2}\)-depleted chloroplasts in the light in the absence of HCO\(\frac{1}{2}\). Thus, bicarbonate action is possible even in the presence of a strong negatively charged membrane surface. We suggest that the uncharged CO\(\frac{1}{2}\) can pass this charged surface by diffusion. Also, Sarojini and Govindjee [24] considered it likely that CO\(\frac{1}{2}\) is the species that diffuses into the membrane.

Furthermore, Stemler [21] suggested that CO_2 is initially taken up by the thylakoid membrane. We propose that after the negative shield has been passed, CO_2 is converted into HCO_3^- which is able to bind to a site below the membrane surface (Fig. 6). This suggestion is in accordance with our conclusion that the rate-limiting step in bicarbonate action is diffusion and that the optimum for the bicarbonate effect is at pH 6.5, where comparable quantities of CO_2 and HCO_3^- are present in the bulk (Fig. 4). However, we cannot exclude the possibility that CO_2 can also bind to the site.

To explain our results we propose a hypothetical model, presented in Fig. 6. According to this model HCO_3^- binds to a site at A and B below the surface of the thylakoid membrane. Only when A and B are both occupied is electron transport between Q and plastoquinone possible. HCO_2^- can bind to the B part of the site, but then electron transport cannot proceed because A is unoccupied. To reach the HCO_3^- -binding site, ions have to pass negative charges at the surface of the membrane and along the path from the surface to the site. In the light, the negatively charged HCO_2^- is not able to leave its binding site. In the dark, the ζ -potential is more positive [23] and, HCO_2^- can leave its site easier

and exchange for HCO₃, thus inducing restoration of electron flow.

Our model also accommodates observations made by Stemler [25] that washing of chloroplasts with silicomolybdate removes HCO3 bound to the site and that DCMU, given prior to the silicomolybdate washing, strongly decreases HCO₃ removal. Stemler concluded that DCMU 'overlays' the bound HCO₃, protecting it from silicomolybdate attack. Furthermore, Barr and Crane [26] have reported that addition of 20 mM HCO₃ results in a full inhibition of the silicomolybdate Hill reaction at pH 8.0. At pH 6.0, however, where the HCO₃/CO₂ ratio is much lower, this amount of added HCO₃ causes only a partial inhibition. This supports our hypothesis that silicomolybdate binds to the same site as HCO₃. Van Rensen and Vermaas [15] showed that DCMU and also the s-triazine herbicide simeton are partial competitive inhibitors of HCO₃ binding. Both compounds decrease the affinity of the thylakoid membrane for HCO₃. Khanna et al. [27] have found that CO₂ depletion causes an altered binding of [14C]atrazine.

It appears that a number of anions can bind to the HCO₃-binding site: HCO₃, HCO₂, silicomolybdate and the herbicide 4,6-dinitro-o-cresol [15].

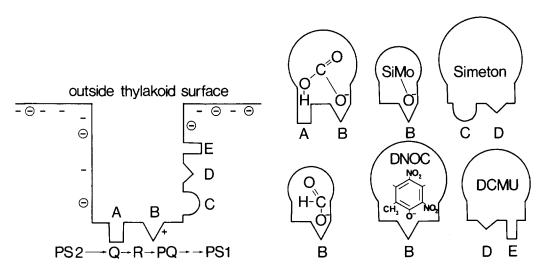


Fig. 6. A hypothetical scheme for the HCO_3^- -binding site. —, charge existing in dark and light; \ominus , charge existing in the light only. B is a nonspecific binding site, to which molecules with $a = C \cdot O^-$ group can bind. Electron transport can only proceed when there is also binding to site A; only HCO_3^- is able to do so. The herbicides, DCMU and simeton, bind to the side of the pathway from the bulk to the HCO_3^- -binding side, thus covering part of the HCO_3^- -binding site. This causes a lowering of the affinity of the site for HCO_3^- . SiMo, silicomolybdate; DNOC, 4,6-dinitro-o-cresol; PS, photosystem.

Thus, the binding site is nonspecific in binding anions, but is very specific in its requirement for HCO_3^- to restore electron transport. It may be that the binding site of these anions and herbicides is located within the Photosystem II shielding protein proposed by Renger [28] and that the shielding protein has partly different binding sites for various classes of Photosystem II inhibiting herbicides, as suggested by Trebst and Draber [29].

Acknowledgements

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References

- 1 Warburg, O. and Krippahl, G. (1958) Z. Naturforsch. 13b, 509-514
- 2 Govindjee, Pulles, M.P.J., Govindjee, R., Van Gorkom, H.J. and Duysens, L.N.M. (1976) Biochim. Biophys. Acta 449, 602-605
- 3 Khanna, R., Govindjee and Wydrzynski, T. (1977) Biochim. Biophys. Acta 462, 208-214
- 4 Siggel, U., Khanna, R., Renger, G. and Govindjee (1977) Biochim. Biophys. Acta 462, 196-207
- 5 Govindjee and Van Rensen, J.J.S. (1978) Biochim. Biophys. Acta 505, 183-213
- 6 Good, N.E. (1963) Plant Physiol. 38, 298-304
- 7 Stemler, A. (1979) Biochim. Biophys. Acta 545, 36-45
- 8 Stemler, A. and Govindjee (1973) Plant Physiol. 52, 119-123
- 9 Vermaas, W.F.J. and Van Rensen, J.J.S. (1981) Proc. 5th Int. Congr. Photosynth., Greece, in the press
- 10 Van Rensen, J.J.S., Wong, D. and Govindjee (1978) Z. Naturforsch. 33c, 413-420

- 11 Van Rensen, J.J.S., Van der Vet, W. and Van Vliet, W.P.A. (1977) Photochem. Photobiol. 25, 579-583
- 12 Renger, G. (1976) FEBS Lett. 69, 225-230
- 13 Itoh, S. and Nishimura, M. (1977) Biochim. Biophys. Acta 460, 381-392
- 14 Van Rensen, J.J.S. and Kramer, H.J.M. (1979) Plant Sci. Lett. 17, 21-27
- 15 Van Rensen, J.J.S. and Vermaas, W.F.J. (1981) Physiol. Plant. 51, 106-110
- 16 Crane, F.L. and Barr, R. (1977) Biochem. Biophys. Res. Commun. 74, 1362-1368
- 17 Schapendonk, A.H.C.M., Hemrika-Wagner, A.M., Theuvenet, A.P.R., Wong Fong Sang, H.W., Vredenberg, W.J. and Kraayenhof, R. (1980) Biochemistry 19, 1922-1927
- 18 Metzner, H. (1975) J. Theor. Biol. 51, 201-231
- 19 Metzner, H., Fischer, K. and Bazlen, O. (1979) Biochim. Biophys. Acta 548, 287-295
- 20 Stemler, A. (1980) Biochim. Biophys. Acta 593, 103– 112
- 21 Stemler, A. (1980) Plant Physiol. 65, 1160-1165
- 22 Stemler, A. (1981) Proc. 5th Int. Congr. Photosynth., Greece, in the press
- 23 Kraayenhof, R. (1977) in Structure and Function of Energy-Transducing Membranes (Van Dam, K. and Van Gelder, B.F., eds.), pp. 223-236, Elsevier, Amsterdam
- 24 Sarojini, G. and Govindjee (1980) Proc. 5th Int. Congr. Photosynth., Greece, Abstr.
- 25 Stemler, A. (1977) Biochim. Biophys. Acta 460, 511-522
- 26 Barr, R. and Crane, F.L. (1980) Biochim. Biophys. Acta 591, 127-134
- 27 Khanna, R., Pfister, K., Keresztes, A., Van Rensen, J.J.S. and Govindjee (1981) Biochim. Biophys. Acta 634, 105-116
- 28 Renger, G. (1976) Biochim. Biophys. Acta 440, 287-300
- 29 Trebst, A. and Draber, W. (1979) in Advances in Pesticide Science, part 2 (Geissbühler, H., ed.), pp. 223-234, Pergamon Press, Oxford